



## Topspin Setup Acquisition: Bio-NMR

Detlef Moskau

Bruker BioSpin



TopSpin

# Setup the Acquisition: Flowchart

# TopSpin Acquisition setup

Data set:

where to define parameters and store spectra

Sample:

temperature, insert, eject, lock, rotate, shim

Standard Parameter sets:

probe independent parameters

Probe dependent parameters:

pulses and power levels

Customize parameters:

spectral windows, number of scans,...

Optimize parameters:

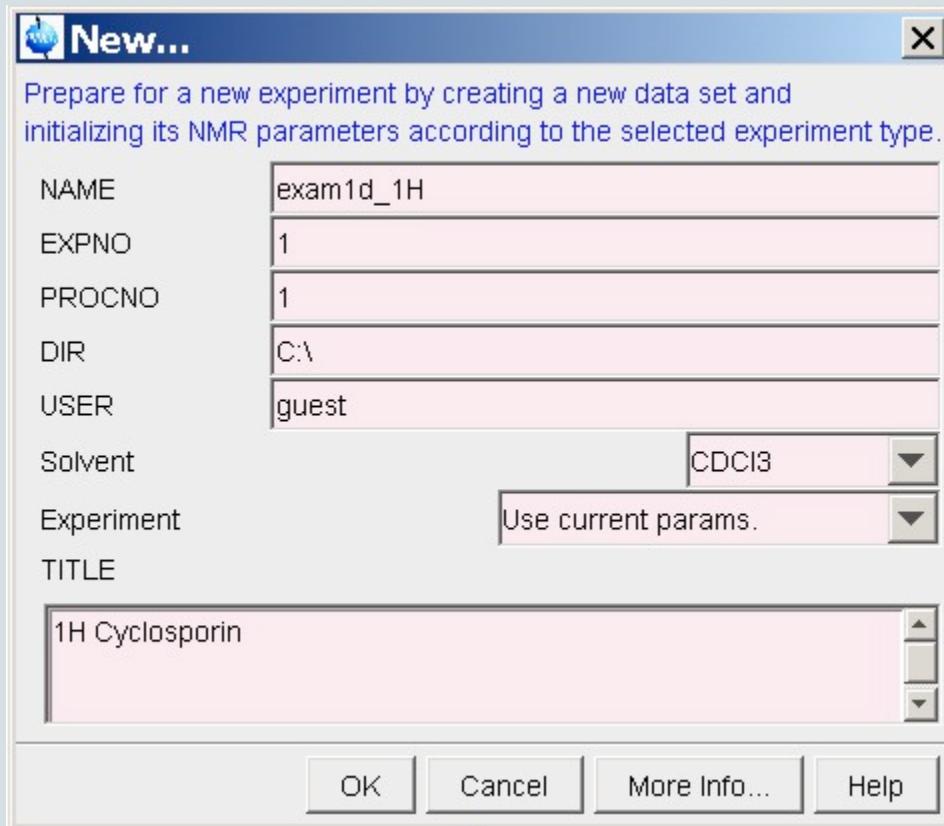
offsets, power levels, receiver gain

Acquisition:

start and stop

# TopSpin Data set commands

Data set: where to define parameters and store spectra  
new, edc



Raw data (FID) is stored under:  
 C:\data\guest\nmr\exam1d\_1H\1

↓      ↓      ↓

USER NAME/PROJECT EXPNO

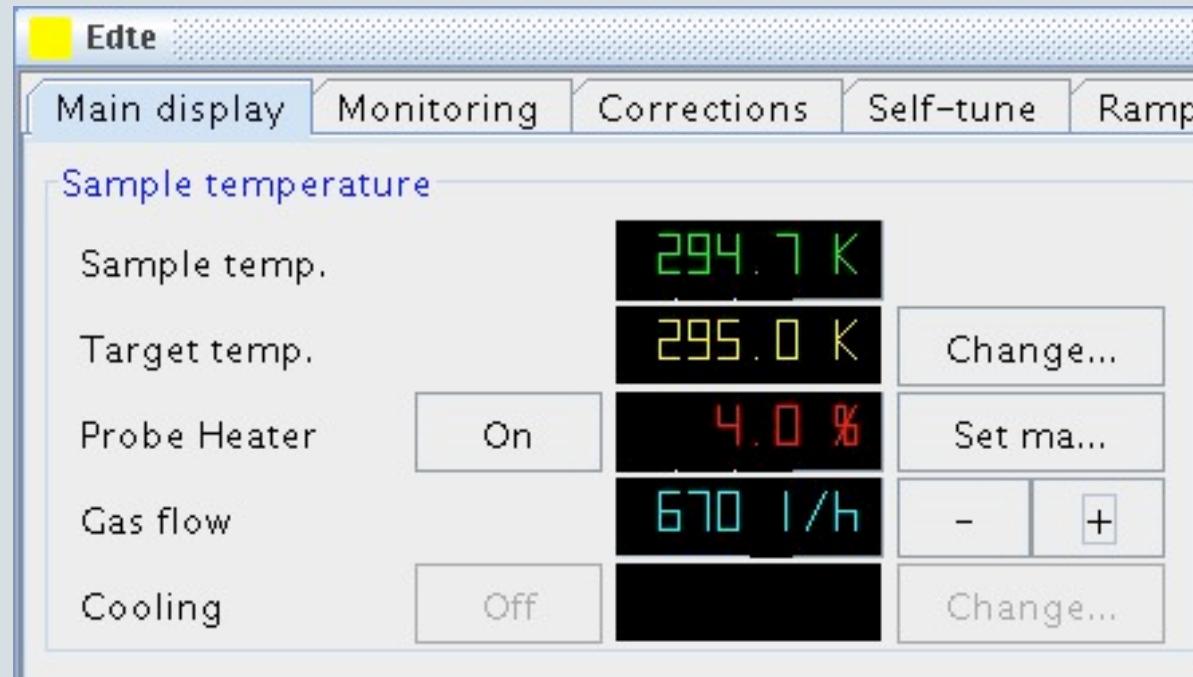
Processed data stored under:  
 C:\data\..\exam1d\_1H\1\pdata\1

↓

PROCNO

## TopSpin Sample

Sample: temperature, insert, eject, lock, rotate, shim  
edte (te and teset)



### Note for CryoProbe:

- Temperature range: 0° – 50° C (temperature has to be calibrated!)
- Gas flow: 670 l/h



## TopSpin Sample

Sample: temperature, insert, eject, lock, rotate, shim  
bsmsdisp: for all functions  
ej, ij: eject, insert sample  
ro: start, stop rotation  
lock: lock  
lock –noauto: lock on solvent with multiple solvent signals (MeOD)

# TopSpin Sample

Sample: temperature, insert, eject, lock, rotate, shim  
lock: for all functions

Solvent	Description
Acetic	acetic acid-d4
Acetone	acetone-d6
C6D6	benzene-d6
CD2Cl2	methylenechloride-d2
CD3CN	acetonitrile-d3
CDCl3	chloroform-d
CH3CN+D2O	HPLC Solvent (Acetonitril/D2O)
CH3OH+D2O	HPLC Solvent (Methanol/D2O)
D2O	deuteriumoxide
DEE	diethylether-d10
Dioxane	dioxane-d8
DME	dimethylether-d6
DMF	dimethylformamide-d7
DMSO	dimethylsulfoxide-d6
EtOD	ethanol-d6
<b>H2O+D2O</b>	<b>90%H2O and 10%D2O</b>
MeOD	methanol-d4
Pyr	pyridine-d5
THF	tetrahydrofurane-d8
Tol	toluene-d8



## TopSpin Sample

Sample: temperature, insert, eject, lock, rotate,  
shim

**topshim:** latest gradient shimming tool  
**gradshim:** alternative gradient shimming  
tool



## TopSpin Experiment definitions

Standard Parameter sets: probe independent parameters

Probe dependent parameters: pulses and power levels



## TopSpin Standard Parameter files

Standard Parameter sets: probe independent parameters

**rpar** (*example: rpar HNCOGP3D all*)

Standard parameter files contain:

Default parameters for an experiment: pulse program, number of scans, spectra windows, time domain data points, window functions for processing etc.

Standard parameter files do **NOT** contain:

Parameters for pulses and power levels



## TopSpin Standard Parameter files

How do I interprete the meaning of the parameter file name?:  
*example: rpar HNCOGP3D all*

### edpul Pulprog.info

;Pulprog.info

;The two-character codes used are the following:

ar	experiment for aromatic residues
at	adiabatic TOCSY
bi	with bird pulse for homonuclear J-decoupling
bp	using bipolar gradients
cc	cross correlation experiment
cp	with composite pulse
ct	constant time
cw	decoupling using cw command
cx	using CLEANEX_PM
dc	decoupling using cpd
.....	



## TopSpin Probe dependent parameters

Probe dependent parameters: pulses and power levels  
**getprosol**

getprosol: load default pulses and power levels according to  
the current **probe** and **solvent**

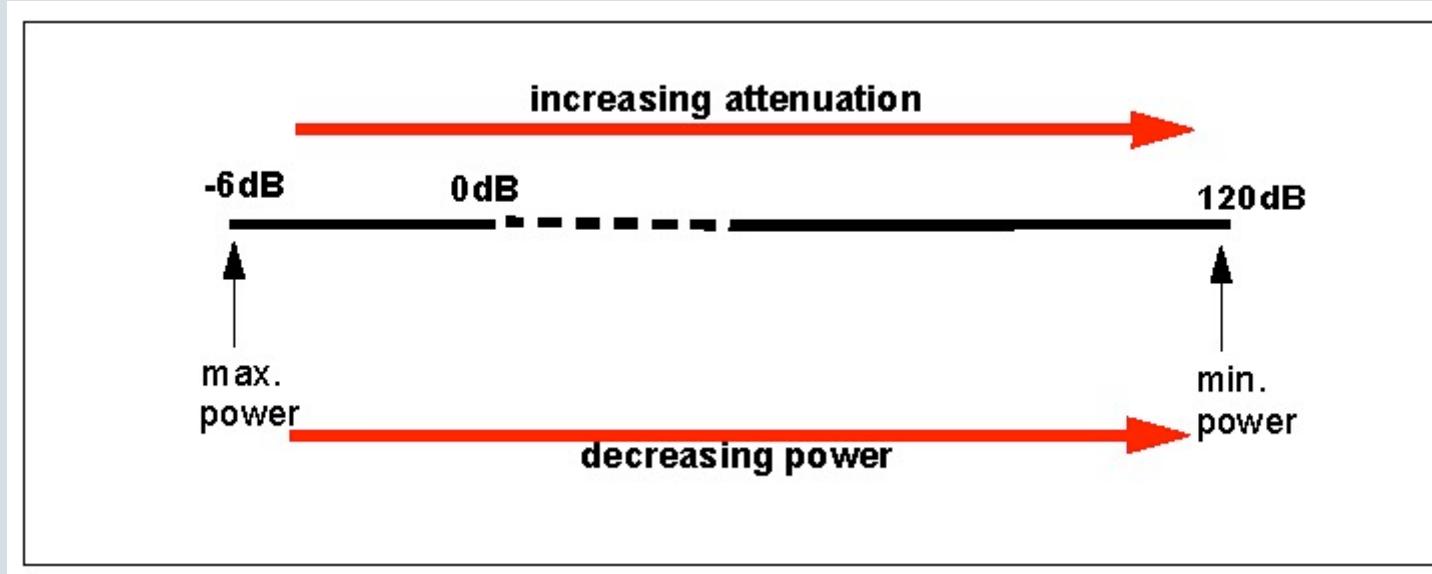
When a pulse does not correspond to the default value, all  
power levels for pulses of that nucleus can be recalculated for  
the current data set:

**EXAMPLE!:** **getprosol 1H 10.5us -2.3db**

for a 1H pulse of 10.5 usec at -2.3 dB

## TopSpin Power level definitions

Power levels are defined as attenuation values in dB:





## TopSpin Customizing parameters

Customize parameters: spectral windows, number of scans,..

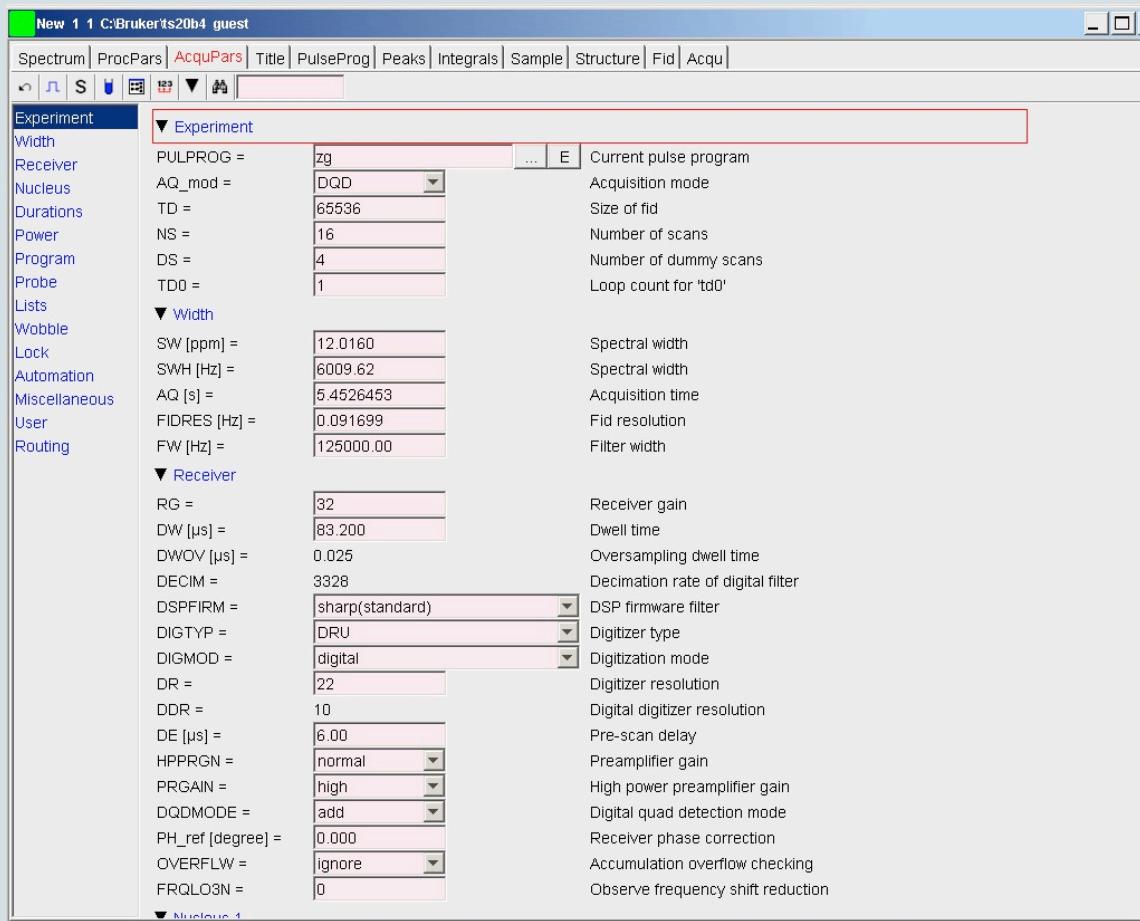
Acquisition:

**eda:** show all parameters

**ased:** show parameters related to current pulse  
program

# TopSpin Customizing parameters

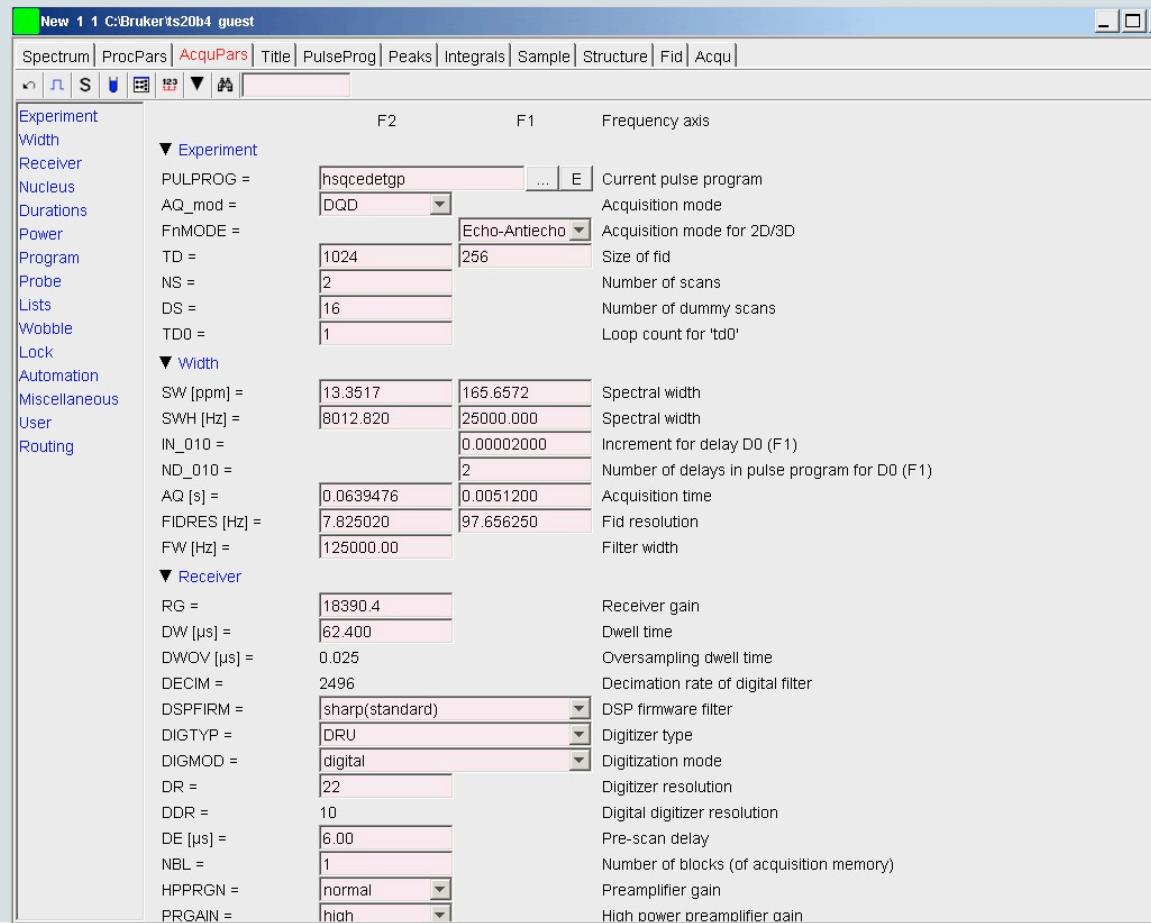
eda: show all parameters



1D-experiments

# TopSpin Customizing parameters

eda: show all parameters



2D-experiments

# TopSpin Customizing parameters

ased: show parameters related to current pulse program

exam1d\_1H 1 1 C:\Bruker\ts20b4 guest

Spectrum | ProcPars | AcquPars | Title | PulseProg | Peaks | Integrals | Sample | Structure | Fid | Acqu |

General

Channel f1

PULPROG =	zg	...	E	Pulse program for acquisition
TD =	65536			Time domain size
NS =	16			Number of scans
DS =	4			Number of dummy scans
SWH [Hz] =	6009.62			Sweep width in Hz
AQ [s] =	5.4526453			Acquisition time
RG =	32			Receiver gain
DW [ $\mu$ s] =	83.200			Dwell time
DE [ $\mu$ s] =	6.00			Pre-scan-delay
D1 [s] =	1.00000000			Relaxation delay; 1-5 * T1
TDO =	1			Dimension of accumulation loop
<b>▼ Channel f1</b>				
NUC1 =	1H	Edit...		Nucleus for channel 1
P1 [ $\mu$ s] =	9.20			F1 channel - high power pulse
PL1 [dB] =	-2.00			F1 channel - power level for pulse (default)
PL1W [W] =	20.30346489			F1 channel - power level for pulse (default)
SFO1 [MHz] =	500.1325007			Frequency of observe channel



## TopSpin Customizing parameters

Tuning and Matching of the probe:

**wobb:** manual mode, no Automatic Tuning and Matching accessory  
(VU: w1, w2, w3 or wobb f1, wobb f2, ...)

**atma:** fully automated tuning and matching, all active channels

**atmm:** manual mode of atma

## TopSpin Customizing parameters

Typical parameters which are customized:

- ns:** number of scans
- ds:** number of dummy scans
- td:** time domain data points
- sw:** spectral window [ppm]
- swh:** spectral window [Hz]
- o1p:** transmitter offset [ppm]
- o1:** transmitter offset [Hz]
- o2p:** decoupler offset [ppm]
- rg:** receiver gain

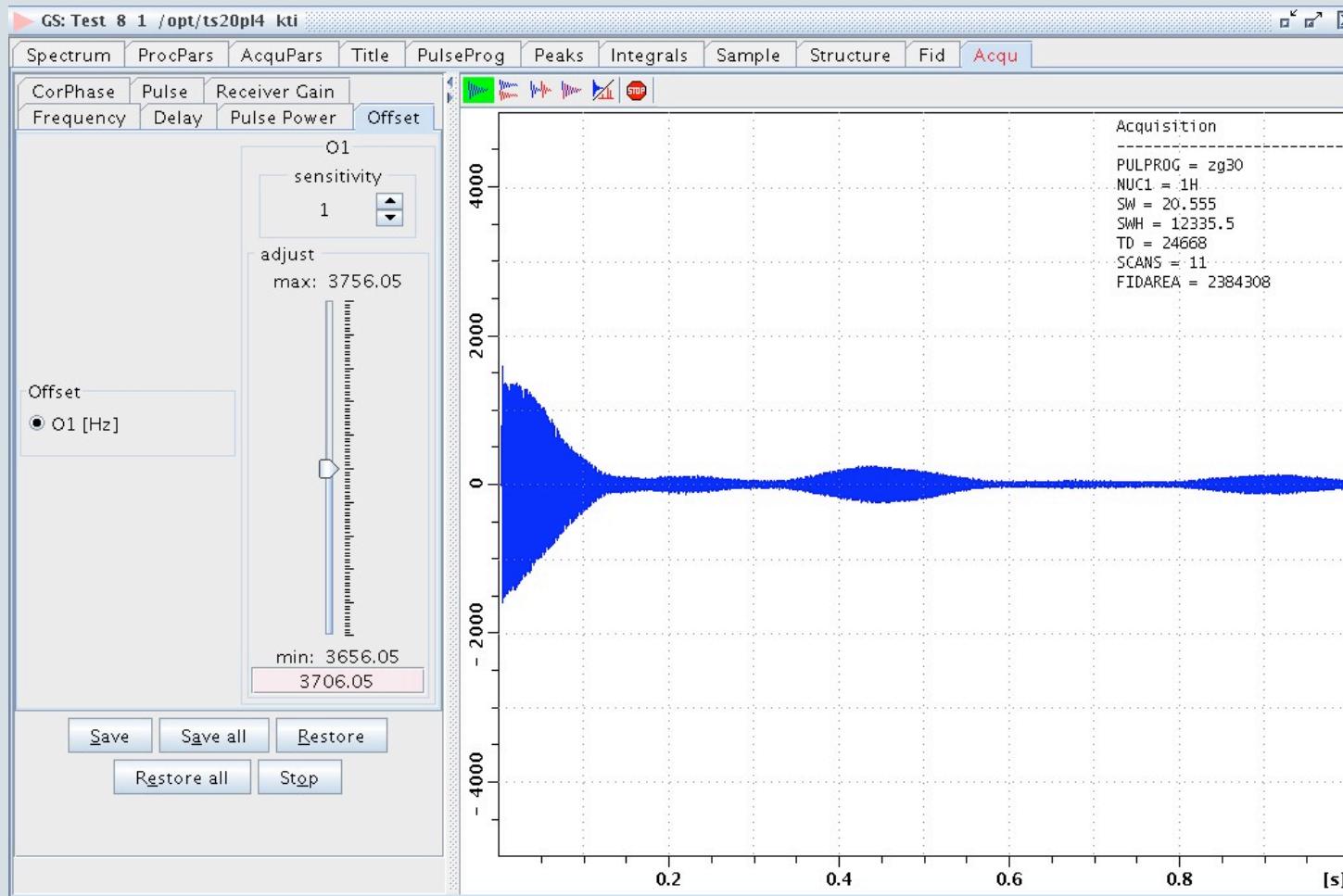


## TopSpin Optimize parameters

Parameters, which need fine tuning (transmitter offset for presaturation, power level for shaped flip-back pulses) can be optimized in the go-setup mode **gs**.

# TopSpin Optimize parameters

go-setup mode gs





# TopSpin Acquisition Start and Stop

## Acquisition: start and stop

### Preparing start of acquisition:

**expt** calculate experiment time

**rga** automatic receiver gain adjustment

### Start acquisition:

**zg** start and overwrite existing FID

**go** start and add to current FID (for 1D)

### Stop acuisition:

**stop** stop immediately. For 1D, FID is lost!

**halt** stop after current scan, FID will be saved

### Transfer 1D-FID to disk:

**tr** required for 1D only



## TopSpin Flowchart commands for acquisition

Data set:	new, edc
Sample:	edte, lock, bsmsdisp, topshim, gradshim rpar
Standard Parameter sets:	
Probe dependent parameters:	getprosol
Customize parameters:	ased, eda
Optimize parameters:	wobb, atma, rg, rga, gs
Acquisition:	expt, zg, stop, halt, tr



## TopSpin Flowchart commands for acquisition

Data set:	new, edc
Sample:	edte, lock, bsmsdisp, topshim, gradshim <b>rpar</b>
Standard Parameter sets:	
Probe dependent parameters:	getprosol
Customize parameters:	eda, ased
Optimize parameters:	wobb, atma, rg, rga, gs
Acquisition:	expt, zg, stop, halt, tr

# Selected list of parameter sets

CBCACONH**GPWG3D**

CBCACONH**GP3D**

CCANH**GP3D**

CCA**CONHGP3D.2**

CCA**CONHGP3D**

CBCANH**GPWG3D**

**CBCANHGP3D**

CCCONHGP3D3

CCCONHGP3D2

CCCONHGP3D1

CCCONHGP3D

CCANHGP3D.2

CCCONHGPWG3D3

CCCONHGPWG3D2

DIPSIHSQCF3GPSI3D

HCACOGPJC3D

HCACOGP3D

HBHANHGPWG3D

HBHANHGP3D

HBHA**CONHGPWG3D**

HBHA**CONHGP3D**

HACAH**B COSYGP3D**

HCC**CONHGPWG3D3**

HCC**CONHGPWG3D2**

HCC**CONHGP3D3**

HCC**CONHGP3D2**

HCC**CONHGP3D1**

HCANNHGP3D

HCA**HBCOSYGP3D**

HCCHE**COSGP3D**

HCCHDIGP3D2

**HCCHDIGP3D HCCH-TOCSY**

HCCHCOSYGP3D

HCCHCOGP3D

HNCACBGPJC3D

**HNCACBGP3D**

HNCACOSYGP3D

HNCACOGPWG3D

HNCACOGP3D

**HNCACBGPWG3D**

HNCAH3D

**HNCAGPWG3D Watergate**

**HNCAGP3D**

HNCAECOSGP3D2

HNCAECOSGP3D

HNCADQZQGP3D

HNCOCACGP3D

**HNCOCACBGPWG3D**

**HNCOCACBGP3D**

HNCOCACAGP3D

HNCAJCGP3D

HNCAHAGP3D

HNCOECSGP3D

HNCOCOGP3D

HNCOCGP3D

HNCOCAGPWG3D

HNCOCAGP3D

HNHBECOSGP3D

**HNHAGP3D**

HNCOGPWG3D

HNCOGPHB3D2

HNCOGPHB3D

**HNCOGP3D**

HNNHDIGP3D

HNHBGP3D

MLEVHSQCETGP3D

MLEVHSQCETF3GP3D

**NA = Nucleic Acid**

NA\_HCCHFWDIECGP3D

NA\_HCCHECGP3D

NA\_GHCCHFWDIGP3D

NA\_HCPDIETGPSI3D

NA\_HCNETGPSISP3D

NA\_HCNETGPSI3D

NA\_HCCNHDIGPWG3D

NA\_HCCHFWDIGP3D

NA\_HCPETGPSI3D

NOESYHSQCF3GP193D

NOESYHSQCETGP3D

NOESYHSQCETF3GP3D

NOESYHSQCGPSM3D

NOESYHSQCGPSISM3D

NOESYHSQCFPF3GPSI3D

**NOESYHSQCF3GPSI3D**

**TR = TROSY**

TRHNCACOGP3D

TRHNCACBGP3D

TRCBCANHGP3D

TRCBCACONHGP3D

TRHNCAGP3D2.2

TRHNCAGP3D.2

**TRHNCAGP3D2 latest version**

TRHNCAGP3D

TRHNCOGP3D

TRHNCOCAGP3D

CCACONHGP2H3D

CCCONHGP2H3D

CCANHGP2H3D

HNCACOGP2H3D

HNCACBGP2H3D

HNCACOGP2H3D.2

**HNCAGP2H3D <sup>2</sup>H-decoupled**

HNCOCAGP2H3D

HNCOCACBGP2H3D

HNCOGP2H3D

TRHNCACBGP2H3D

TRHNCAGP2H3D2

TRHNCAGP2H3D

TRHNCACOGP2H3D

TRHNCOGP2H3D

TRHNCOCAGP2H3D

TRHNCOCACBGP3D

TRHNCOCACBGP2H3D



## TopSpin Standard Parameter files

How do I interprete the meaning of the parameter file name?:  
*example: rpar HNCOGP3D all*

### edpul Pulprog.info

;Pulprog.info

;The two-character codes used are the following:

ar	experiment for aromatic residues
at	adiabatic TOCSY
bi	with bird pulse for homonuclear J-decoupling
bp	using bipolar gradients
cc	cross correlation experiment
cp	with composite pulse
ct	constant time
cw	decoupling using cw command
cx	using CLEANEX_PM
dc	decoupling using cpd
.....	



## Example: rpar HNCA\*

**HNCAGP3D** gradient selected

**HNCAGPWG3D** watergate version

**HNCAGP2H3D**  $^2\text{H}$ -decoupled /  $^1\text{H}$ -suppression by  $180^\circ$ -pulses

**TRHNCAGP3D2** TROSY /  $^1\text{H}$ -suppression by  $180^\circ$ -pulses

**TRHNCAETGP3D** TROSY / echo-antiecho /  $^1\text{H}$ -suppression by  $180^\circ$ -pulses  
(JMR 144 (2000) 123)

**TRHNCAGP2H3D** TROSY /  $^2\text{H}$ -decoupled / for 100% deuterated proteins

**TRHNCAGP2H3D2** TROSY /  $^2\text{H}$ -decoupled / for partially deuterated proteins  
/  $^1\text{H}$ -suppression by  $180^\circ$ -pulses

# Other triple resonance parameter sets

**HNCOCAGP3D**

**HNCOCAGP**WG3D****

**HNCOCAGP**2H3D****

**TRHNCOCAGP3D**

**TRHNCOCAGP**2H3D****

**HNCOCACBGP3D** „transfer“

**HNCOCACBGP**WG3D****

**HNCOCACBGP**2H3D****

**TRHNCOCACBGP3D**

**TRHNCOCACB**ETGP3D**** echo-antiecho

**TRHNCOCACBGP**2H3D**** <sup>2</sup>H-decoupled

**HNCACBGP3D** „out&back“/ for larger proteins

**HNCACBGP**WG3D****

**HNCACB**IGP**WG3D****** intra-residue only

**HNCACBGP**2H3D****

**TRHNCACBGP3D**

**TRHNCACB**IGP3D**** intra-residue only

**TRHNCACBGP**2H3D****

**HAHCONHGP3D**

**HAHCONHGP**WG3D****

**TRHAHCONHGP3D**

**HAHBNHGP3D**

**HAHBNHGP**WG3D****

**TRHAHBNHGP3D**

**HNCOGP3D**

**HNCOGP**WG3D**** watergate

**TRHNCOGP3D** TROSY

**HNCACOGP3D**

**HNCACOGP**WG3D****

**HNCACOGP**2H3D**** <sup>2</sup>H-decoupled

**TRHNCACOGP3D**

**TRHNCACOGP**2H3D****

**CBCACONHGP3D** „transfer“ / for smaller proteins

**CBCACONHGP**WG3D****

**TRBCACONHGP3D**

**CBCANHGP3D** „transfer“ / for smaller proteins

**CBCANHGP**WG3D****

**TRBCANHGP3D**

**TRBCANH**ETGP3D****

**SEQTRBCANHGP3D** sequential TROSY

**HCCCONHGP3D2** H(CCCO)NH

**HCCCONHGP**WG3D2****

**HCCCONHGP3D3** (H)CC(CO)NH

**HCCCONHGP**WG3D3****

**CCCONHGP**2H3D**** <sup>13</sup>C-start

## Other useful parameter sets

### <sup>13</sup>C-correlated experiments

**HCCHDI**GP3D

**HCCH**-TOCSY

**HCCHDI**GP3D2

(**H**)CCH-TOCSY

**HCCHCO**GP3D

**HCCH**-COSY

**NOESYHSQC**ETGP3D

**3D-NOESY-HSQC**

### <sup>15</sup>N-correlated experiments

**NOESYHSQC**F3GPSI3D

**15N-NOESY-HSQC** sensitivity enhanced

**NOESYHSQC**F3GP193D

**15N-NOESY-HSQC** with 3919 watergate

**NOESYTZ**GP3D

**15N-NOESY-ZQ-TROSY** for large proteins

## Other useful parameter sets

### 2D Homonuclear-correlated experiments

**NOESYPHPR**

**NOESY, phase sensitive, presat**

**NOESYGPPH19SW**

**NOESY, phase sensitive, 3919 WATERGATE**

**MLEVPHPR**

**TOCSY, phase sensitive, presat**

**MLEVGPPH19SW**

**TOCSY, phase sensitive, 3919 WATERGATE**

**DIPSI2GPPH19**

**TOCSY, phase sensitive, 3919 WATERGATE,  
DIPSY2 spin lock**

### Where is the parameter file for a NOESY with Excitation Sculpting?

Not all versions of an experiment are covered by parameter files

Start from a related experiment and:

- change the pulse program: **pulprog noesyesgpph instead noesyphpr**
- check corresponding parameters with **ased**



## Other useful parameter sets

### 2D Heteronuclear-correlated experiments

<b>13C:</b>	<b>HSQCETGP</b>	<b>HSQC, echo-antiecho</b>
	<b>HSQCETGPSISP</b>	<b>HSQC, echo-antiecho, sensitivity improved, adiabatic <math>^{13}\text{C}</math> inversion</b>
	<b>HSQCETGPSISP.2</b>	<b>HSQC, echo-antiecho, sensitivity improved, adiabatic <math>^{13}\text{C}</math> inversion and refocussing</b>
<b>15N;</b>	<b>HSQCETF3GP</b>	<b>HSQC, echo-antiecho</b>
	<b>HSQCETF3GPSI</b>	<b>HSQC, echo-antiecho, sensitivity improved</b>



# Quick Guide

## Let's run a 3D CBCACONH with WATERGATE

edc

rpar CBCACONHGPWG3D all

lock

atma f1

topshim

pulseCAL

getprosol 1H x us y db

ased

eda

rg

expt

zg

new data set

read parameter file

lock on solvent

tune and match 1H channel automatically

gradient shimming

determine 1H pulse automatically

set all pulses, recalculate all 1H pulses

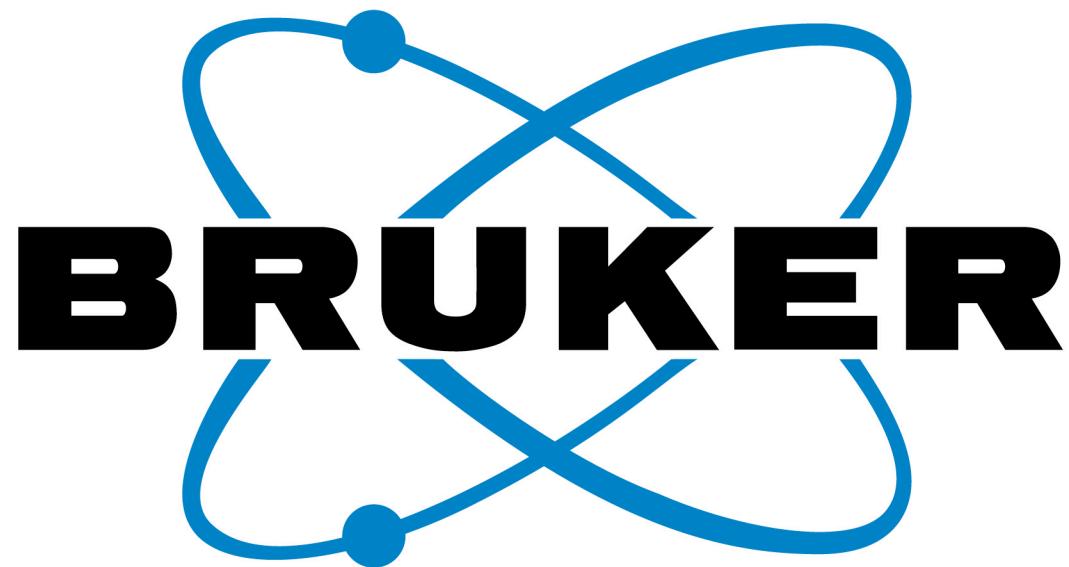
all pulse parameters and delay correct?

check NS, spectral windows and number of increments

typically = 200

check total experiment time

start the acquisition



[www.bruker-biospin.com](http://www.bruker-biospin.com)

Bruker BioSpin